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PATENT

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: Roberto Malinow *et al*	)	Examiner: Not yet assigned
Serial No.: Not yet assigned	)	Art Unit: Not yet assigned
Filed: July 14, 1999	)	**UTILITY PATENT APPLICATION TRANSMITTAL (37 C.F.R. § 1.53(b))**
For: **Diagnostic Methods for Drug Screening for Alzheimer's Disease**	)	

BOX PATENT APPLICATION
Commissioner of Patents and Trademarks
Washington, D.C. 20231

Sir:

This is a request for filing a patent application under 37 C.F.R. § 1.53(b) in the name of inventors: Roberto Malinow, Sahid Zaman, Sangram S. Sisodia, David R. Borchelt, and Michael K. Lee

For: **Diagnostic Methods for Drug Screening for Alzheimer's Disease**

This application is a [X] Continuation [ ] Divisional [] Continuation-in-part of prior Application No.: 09/193,221 filed November 16, 1998, from which priority under 35 U.S.C. § 120 is claimed.

Application Elements:

14 Pages of Specification, Claims and Abstract

4 Sheets of **formal** Drawings

CERTIFICATE OF EXPRESS MAILING

"Express Mail" Label No.: EL288327221US

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I hereby certify under 37 C.F.R. 1.10 that this correspondence is being deposited with the United States Postal Service as "Express Mail Post Office to Addressee" with sufficient postage on the date indicated above and is addressed to the Commissioner of Patents and Trademarks, Washington, D.C. 20231.

(Signature)

ANDREA JOHNSON

(Printed Name)

[X] Declaration

[ ] Unexecuted Combined Inventor Declaration and Power of Attorney

[X] Copy from prior application (37 CFR 1.63(d) for a continuation or divisional).

The entire disclosure of the prior application from which a copy of the declaration is herein supplied is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

[ ] Deletion of inventors Signed statement attached deleting inventor(s) named in the prior application, *see* CFR 1.63(d)(2) and 1.33(b).

Accompanying Application Parts:

[ ] Assignment and Assignment Recordation Cover Sheeet (recording fee of $40.00 enclosed)
[ ] Power of Attorney
[ ] 37 CFR 3.73(b) Statement by Assignee
[ ] Information Disclosure Statement with Form 1449
[ ] Copies of IDS Citations
[ ] Preliminary Amendment
[X] Return Receipt Postcard
[X] Small Entity Statement(s)
[X] Statement filed in prior application.
Status still proper and desired.
[ ] Other:
[ ] A sequence listing.
[ ] paper copy.
[ ] computer readable copy.
[ ] Statement in Compliance with Requirements for Patent Applications Containing Nucleotide and/or Amino Acid Sequence.

Claim For Foreign Priority

[ ] Priority of _____ Application No. _____ filed on _____ is claimed under 35 U.S.C. § 119
[ ] The certified copy has been filed in prior application U.S. Application No.09/193,221
[ ] the certified copy will follow.

Extension of Time for Prior Pending Application

[ ] A Petition for Extension of Time is being concurrently filed in the prior pending application. A copy of the Petition for Extension of Time is attached.

Amendments

[X] Amend the specification by inserting before the first line the sentence: "This is a [X] Continuation [ ] Continuation-in-part [ ] Divisional application of copending prior

[X] Application No. 09/193,221 filed on November 16, 1999.

[ ] International Application _____ filed on _____, which designated the United States, disclosure of which is incorporated herein by reference."

[ ] Cancel in this application original claims _____ of the prior application before calculating the filing fee

Fee Calculation (37 CFR § 1.16):

	(Col. 1)	(Col. 2)	SMALL ENTITY		OTHER THAN A SMALL ENTITY	
FOR:	NO. FILED	NO. EXTRA	RATE	FEE	RATE	FEE
Basic Fee			$380	$380	$760	$
Total Claims	12	0	$ 9	$	$ 18	$
Indep Claims	4	1	$ 39	$ 39	$ 78	$
[] Multiple Dependent Claims			$130	$	$260	$
Total Filing Fee:				$419		$

TOTAL FEES: $419.00

[X] A check including the amount of the above-indicated TOTAL FEES is attached.

[ ] Please charge Deposit Account No.18-0020 in the amount of $_____.

[] A check in the amount of $_____ is attached.

[ ] No fee is required.

[X] Conditional Petition for Extension of Time: An extension of time is requested in the present and/or the above-referenced parent application to provide for timely filing if an

extension of time is still required after all papers filed with this transmittal have been considered.

[X] The Commissioner is hereby authorized to charge any underpayment of the following fees associated with this communication, including any necessary fees for extension of time, or credit any overpayment to Deposit Account No. 18-0020.

[X] Any filing fees under 37 CFR 1.16 including fees for the presentation of extra claims.

[X] Any parent application processing fees under 37 CFR 1.17.

[X] A **duplicate** copy of this sheet is attached for accounting purposes.

Respectfully submitted,

Dated: July 14, 1999

By: _____
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Enclosures

Applicant or Patentee: Malinow, et al. Attorney's Docket No.: A-67162/BIR
Serial or Patent No.: 09/193,221
Filed or Issued: November 16, 1998
For: **DIAGNOSTIC METHODS FOR DRUG SCREENING FOR ALZHEIMER'S DISEASE**

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS (37 CFR 1.9(f) and 1.27(d)) - NONPROFIT ORGANIZATION

I hereby declare that I am an official empowered to act on behalf of the nonprofit organization identified below:

NAME OF ORGANIZATION COLD SPRING HARBOR LABORATORY
ADDRESS OF ORGANIZATION 1 BUNGTOWN ROAD, COLD SPRING HARBOR, NEW YORK 11724

TYPE OF ORGANIZATION

[ ] UNIVERSITY OR OTHER INSTITUTION OF HIGHER EDUCATION
[X] TAX EXEMPT UNDER INTERNAL REVENUE SERVICE CODE (26 USC 501(a) and 501(c)(3))
[X] NONPROFIT SCIENTIFIC OR EDUCATIONAL UNDER STATUTE OF STATE OF THE UNITED STATES OF AMERICA (NAME OF STATE New York) (CITATION OF STATUTE 501(c)(3))
[ ] WOULD QUALIFY AS NONPROFIT SCIENTIFIC OR EDUCATIONAL UNDER STATUTE OF STATE OF THE UNITED STATES OF AMERICA IF LOCATED IN THE UNITED STATES OF AMERICA (NAME OF STATE _____) (CITATION OF STATUTE _____)

I hereby declare that the nonprofit organization identified above qualifies as a nonprofit organization as defined in 37 CFR 1.9(e) for purposes of paying reduced fees under Section 41(a) or (b) of Title 35, United States Code, with regard to the invention entitled **DIAGNOSTIC METHODS FOR DRUG SCREENING FOR ALZHEIMER'S DISEASE by inventors** Roberto Malinow, Shahin Zaman, Sangram S. Sisodia, David R. Borchelt, Michael K. Lee, described in

[ ] the specification filed herewith
[X] application serial no. 09/193,221, filed November 16, 1998
[ ] patent no. _____, issued _____

I hereby declare that rights under contract or law have been conveyed to and remain with the nonprofit organization having rights to the invention as listed below* and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 CFR 1.9(c) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e). *NOTE: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

NAME: _____
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[ ] Individual [ ] Small Business Concern [ ] Nonprofit Organization

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING John Maroney
TITLE IN ORGANIZATION Asst. Administrative Director
ADDRESS OF PERSON SIGNING 1 BUNGTOWN ROAD, COLD SPRING HARBOR, NEW YORK 11724
SIGNATURE _____ DATE 12/3/98

DIAGNOSTIC METHODS FOR DRUG SCREENING FOR ALZHEIMER'S DISEASE

Technical Field

The field of this invention is the screening of drugs for treatment of Alzheimer's and related neurodegenerative diseases.

Background

Alzheimer's disease (AD) is the most common cause of dementia in the elderly. Mutations in the amyloid precursor protein gene (*APP*) and presenilins (1 and 2; *PS1* and *PS2*) cause autosomal dominant, early-onset forms of AD and account for ~1% and ~50% of inherited cases, respectively. Polymorphisms in the *apoE4* and *α-2 macroglobulin* genes are associated with increased risk in individuals over 60 years of age.

The presenilins are polytopic membrane proteins expressed in the endoplasmic reticulum, Golgi complex in dendrites (close to dendritic spines) and axon terminals in neurons. The PS1 holoprotein is subject to endoproteolysis; the resulting N– and C-terminus fragments bind to each other at stoichiometric levels and/or other proteins, such as γ-catenin. The levels of the fragments are very tightly regulated and overexpression studies show little changes in the relative amounts of accumulated fragments.

The normal biological function(s) of presenilins are not well understood although they have been shown to play a major role in the embryonic development of the axial skeleton and cerebral vasculature. The inheritance pattern in humans carrying mutant presenilin genes suggests a gain-of-function. Several cellular effects of mutant presenilins have been documented that may be relevant to the pathophysiology of AD. First, in cultured cells and transgenic animals expression of

mutant presenilins lead to the elevated production of αβ42(43) peptides that are deposited early and selectively in amyloid plaques in AD. The over-production of Aβ42(43) is most pronounced in cells expressing a PS-1 mutation lacking exon 9 (Δ9). Secondly, cells expressing mutant presenilins also have aberrant calcium homeostasis; PC12 cells expressing mutant PS1 stimulated with agonists that activate Ca^{2+} efflux from intracellular stores, exhibit larger calcium transients than cells expressing wild-type (wt) PS1.

The clinical hallmark of early AD is a disruption of memory processes. The hippocampus, which is prominently involved in the formation of memory, is affected early in the disease and shows the characteristic histopathological changes of AD, namely senile (amyloid) plaques and neurofibrillary tangles. A loss of synapses is also apparent in the hippocampus early in the disease. As the disease progresses, neuronal death in the hippocampus increases (see review by Price et al., 1998 Annu. Rev. Neurosci. 21:479-505).

Hippocampal slices have been effectively used to examine synaptic transmission and plasticity *in vitro*. Stimulation of CA1 *striatum radiatum* afferent pathways produces a mixed excitatory and inhibitory synaptic response in pyramidal neurons. Brief repetitive stimulation generates a short and long-term potentiation (STP: ~20 min and LTP:>30 min) of excitatory transmission that have been proposed as cellular correlates of some forms of learning. STP and LTP also share several underlying mechanisms with glutamate-mediated neuron death (excitotoxicity, see reviews by Obrenovitch et al., 1997 J. Prog. Neurobiol. 51:39-87 and Choi, 1992 J. Neurobiol. 9:1261-1276). All are believed to require synaptically induced postsynaptic depolarization, activation of NMDA receptors and a rise in intracellular calcium concentration. As membrane depolarization is a critical requirement for STP and LTP, these phenomena are sensitive to pharmacological manipulations of fast inhibitory pathways via the $GABA_A$ receptor. In the CA1 region of the hippocampus, for example, a $GABA_A$ antagonist leads to more LTP, while $GABA_A$-potentiating benzodiazepines can reduce LTP. $GABA_A$ agonists can also decrease glutamate-induced excitotoxicity.

Based on the present understanding of the etiology of AD and the neuronal mechanisms associated with AD and memory, there is a need for a diagnostic method for evaluating the potential of drugs for the treatment of AD, both prophylactically and therapeutically.

Brief description of relevant art

The effect of benzodiazepines in decreasing the incidence of AD has been described by Fastbom et al., 1998 Alzheimer Dis. Assoc. Disord. 12:14-17. Mechanisms associated with learning, excitatory transmission and the involvement of $GABA_A$ receptor are described by Bliss et al., 1993 Nature 361:31-39; Wigstrom et al., 1986 J. Physiol. (Paris) 81:228-236; Evans, et al., 1996 Neuropharmacology 35:347-357; Ohkuma et al. 1994 Jpn. J. Pharmacol. 64:125-128; and Muir et al., 1996 J. Cereb. Blood Flow Metab. 16:1211-1218. Biological functions and cellular effects of presenilins are described in Shen et al., 1997 Cell 89:629-639; Wong et al., 1997 Nature 387:288-292; Lee et al., 1997 Nat. Med. 3:756-760; and Borchelt et al., 1996 Neuron 17:1005-1013. Guo et al., 1996 Neuroreport. 8:379-383 report that cells having mutant presenilins have aberrant calcium homeostasis.

SUMMARY OF THE INVENTION

Mutant presenilin comprising hippocampal cells are employed in an assay for screening drugs for the treatment of Alzheimer's disease. Tissue samples from the hippocampus having a presenilin mutation are subjected to tetanic stimulation in the presence of a candidate drug and cellular plasticity is determined, as compared to the presence of a control. The measured outcome is reduction of aberrant signaling.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 shows graphs of a study of basal transmission and paired-pulse facilitation (PPF). Fig. 1a, top, graphs are examples of field responses in hippocampal slice CA1 region evoked by delivery of increasing intensity stimuli for wild-type ("wt")

and Δ9 mutant animals (averages of 5 each). Fig. 1a bottom is the input-output plot of basal transmission in mutant (Δ) and wt animals (O) obtained from responses evoked as in 1a (top). The plot includes data from 4 wt and 6 mutant slices. Best fit line to each group (linear regression) shows slopes that are not significantly different ($p>0.05$). Scale bars: 10ms and 0.2 mV. Fig. 1b top shows examples of responses to paired stimuli (50 ms inter-stimulus interval, averages of 10 each). Fig. 1b bottom is a plot of percent potentiation versus inter-pulse interval for mutant (Δ, n=6 slices) and wt (O, n=6 slices) animals. Values are not significantly different ($p>0.05$). Scale bars: 50 ms and 0.f1 mV.;

Fig. 2 shows graphs of epsp response to tetani in the presence or absence of agents which affect the $GABA_A$ receptor to evaluate the role of $GABA_A$ in the presence or absence of agents which affect the $GABA_A$ receptor. Fig. 2a graphs are based on experiments with $GABA_A$ transmission intact. On the left, examples of epsp response (averages of 10 each) immediately before and 20 min after a tetanus superimposed for wt and mutant animals. Scale bars: 0.2 mV and 20 ms apply to a and b. On the right, plot of mean epsp slope (±SEM) normalized to values before tetanic stimulus (time 0). For each slice transmission two independent pathways were monitored. Tetanized pathway showed greater enhancement in slices from mutant (open triangles, n=8 slices) than wt (open circles, n=10 slices) animals. Control pathways (mutant, closed triangles; wt closed circles) remain unchanged. Tetanus consisted of 100 stimuli delivered over 1 sec (100 Hz). The potentiation at the time points of 5, 10, 15, 20, 25 and 30 min was: 1.36±0.060, 1.26±0.058, 1.27±0.054, 1.24±0.058, 1.22±0.063 and 1.21±0.069, respectively for the wt. For the same time points for the mutant these values were: 1.67±0.065, 1.52±0.054, 1.54±0.072, 1.56±0.075, 1.51±0.089 and 1.54±0.090, respectively. At these time points, thre was a statistically significant difference between the two sets of data points (<0.05). Abbreviation "-PTX", no picrotoxin in the bath. Fig. 2b shows results with $GABA_A$ transmission blocked with 100μM picrotoxin. On the left, are examples of epsp responses (average of 10 each) immediately before and 20 min after a tetanus superimposed for wt (top) and mutant (bottom) animals. On the right, a plot of mean epsp slope±SEM normalized to values before tetanic stimulus

(time 0). For these experiments, control pathways were monitored for only 30 min after tetanus. Tetanized pathways showed similar enhancement in slices from mutant (open triangle, n=13 slices) and wt (open circles, n=18 slices) animals. Control pathways (mutant, closed triangles, wt closed circles) remain unchanged. Tetanus consisted of 25 pulses given as groups of 5 pulses at 100 Hz every 10 s, 5 times. This tetanus was weaker than in Fig. 2a to obviate possible differences between wt and mutant induction. Abbreviation, "+PTX", with picrotoxin in bath. Fig. 2c shows the results of experiments in the presence of N-methyl-D-aspartate (NMDA)-receptor blockade with AP5. Plot of mean epsp slope±SEM normalized to values before tetanic stimulus (time 0).;

Fig. 3 shows the effect of flunitrazepam on LTP. Fig. 3a is a graph of the means±SEM of normalized epsp responses in the absence of drug plotted against time: wt (•, n=7 slices), Δ9 mutation (▲, n=12 slices). There was a significantly greater amount of potentiation in the mutant at the time points of 5, 10 and 15 min post-tetanus. At 20 min, the difference in potentiation became insignificant. The tetanus (delivered at time 0) was 100 pulses given for 1s (100 Hz) every 20s 3x in succession. Control pathways (receiving no tetanus) remain unchanged. Abbreviation, "-FLU", no flunitrazepam in bath. Fig. 3b was as in Fig. 3a but in the presence of flunitrazepam in the bathing medium; (•, n=11 slices) and Δ9 mutation (▲, n=8 slices). There was no statistically significant difference between the two groups at the above time points post-tetanus. Control pathways (receiving no tetanus) remained unchanged. Abbreviation, "+FLU", flunitrazepam present in bath. Fig. 3c is the same data as Figs. 3a and 3b comparing potentiation in mutants (+FLU) with the wt (-FLU) to show the suppression of the potentiation to almost the wt levels. Fig. 3d are histograms from the data of Figs. 4a and 4b showing potentiation at various times post-tetanus. All groups were compared with each other, only statistically significantly different pairs ($p<0.05$) are shown by the lines. These comparisons were calculated for 5 min, 10 min (histogram not shown) and 15 min. These time points gave identical statistical results for pairwise comparisons as in the 5 min case. For the 20 min time point, however, the W vs. M comparison

was not significant but the other two pairwise group comparisons were significant at $p<0.05$. W=wild type, M=mutant, Wf=wt +FLU and Mf=mutant +FLU.;

Fig. 4 are graphs of the effect of agents on the $GABA_A$ receptor-mediated transmission in the mutant and wt cells using whole-cell patch-clamping. Fig. 4a graphs the evoked synaptic response (averaged up to 20 each) from whole-cell patch-clamped neurons. Outward current recorded at 0mV is completely blocked by 100μ PTX, a $GABA_A$ receptor antagonist. NBQX blocked some of the outward current (not shown) indicating some di-synaptic inhibition. At the holding potential of -60mV, the inward current is completely blocked with 2μM NBQX, the AMPA (glutamate subtype) receptor blockade. Scale bar: 25 pA and 20ms. Fig. 4b graphs the evoked synaptic responses (averaged up to 15 each) recorded at holding potentials of 0mV and -60mV with the stimulating electrode placed in stratum radiatum (top) at site 1, ~50μ from the recording electrode (middle) at site 2, ~250μm from the recording electrode and (bottom) at site two with the stimulus intensity increased ~3 -fold. Scale bars: top, 50pA; middle, 100 pA; bottom, 50 pA; time scale, as in Fig. 4a. Fig. 4c, left, examples of averaged (up to 15 each) traces from patch-clamp whole-cell recordings in wt and Δ9 mutation at -60mV (glutamate currents) and at 0mV ($GABA_A$ currents). Scale bar: 40pA and 20ms. On the right, peak amplitude of response ratios (measured at holding potentials of -60mV and 0mV, respectively) from cells in individual slices (n=9 slices each). The means±SEM are also shown and superimposed using the filled symbols. The ratios are significantly greater in the mutant than in the wt ($p<0.05$, t-test).

Fig. 5 graphs the results from the effects of AP5-sensitive potentials during tetanus. Fig. 5a shows normalized traces of field potential response to four consecutive (every 10ms) stimuli, before (the larger response) and after (the intermediate response) the application of the specific NMDA-receptor antagonist, AP5. The differences between the two responses at each time point are also shown. The responses were normalized to the area up to the peak of the first response (which is mostly due to non-NMDA receptor activation). Scale bar: 10ms. Fig. 5b shows the averaged differences of areas under the four response curves, before and after AP5

application in individual (wt and Δ9 mutant) slices to show the effect of tetanus on the NMDA (or AP5)-sensitive component. The means±SEM of all slices are also shown in filled symbols. Although the mean AP5-sensitive potentials were smaller in the mutants (despite manifesting a greater potentiation (see Fig. 2), there was no statistically significant difference between the two groups.

DETAILED DESCRIPTION OF THE INVENTION

A method for screening drugs is provided for determining their potential for the treatment of Alzheimer's disease (AD). The effect of agents on changes in plasticity of mutant cells is related to their ability to treat AD. It is found that cells with presenilin mutations, particularly PS-1, can be used in a battery of tests to evaluate plasticity of cells to tetani, where restoring wild-type behavior indicates potential use as a therapeutic.

Mammalian species may be used as a source of mutant hippocampal tissue. Any mutant which provides the desired enhanced synaptic potentiation upon tetanic stimuli in the same manner as observed with a PS-1 mutation may be employed. This can be achieved in a variety of ways of varying convenience. A transgenic mammalian host can be employed where a mutated presenilin gene is introduced, where it acts as an autosomal dominant allele. Alternatively, one may provide a transgenic host, where presenilin antisense is transcribed from an inducible promoter. Also, one can infect cells or tissue with viruses which provide such genetic capability as described above. In some situations, transformed or otherwise immortalized hippocampal cells may be employed for genetic modification. Other techniques may also be used to provide the desired mutant. The mutation is in a presenilin gene, particularly PS-1. While any mammal may be used as the source of the tissue, for convenience murine species, rats and mice, may be employed, although primates other than humans, or domestic animals, such as porcine, feline, canine, lagomorpha, etc. may also find use. Lee et al., 1997 Nat. Med. 3:756-760, describes hyperaccumulation of FAD-linked presenilin variants in vivo.

In carrying the assay out, there may be an interest in first determining synaptic transmission and plasticity in hippocampal slices of wild-type and mutant hosts. Synaptic transmission is elicited by delivering stimuli of different intensities to afferent pathways. Input-output curves are generated by plotting the slopes of excitatory postsynaptic potentials (epsp) versus fiber volley amplitude (a measure of the number of presynaptic fibers activated). Appropriately, no significant difference should be observed between wt and mutant tissue for use in the assay.

While desirably, one may have wild-type hippocampal cells matched to the mutant cells, by having substantially no genetic difference affecting the assay, as a control, such control is not essential. By knowing the response of the wild-type cells to tetanic stimuli under the conditions of the assay, one can compare the results of the mutant cells with the known standard results. However, it will usually be desirable to have wild-type matched hippocampal cells to ensure that the observed results with the mutant have a direct comparison under the conditions of the assay. The control may be performed with and/or without the candidate drug to provide a comparison with the results from the mutant cells. In addition, one may have a comparison as to the effect of a known drug having a known activity on the mutant cells under the conditions of the assay. In this way one can directly compare the activity of the candidate drug to a known drug, as well as the activity of the candidate drug on wild-type cells in relation to the synaptic potential response to tetani. The assay is usually carried out over an extended period of time taking readings at different time points and determining the potentiation. Normally, the $GABA_A$ transmission by the cells will be intact.

The effect of tetanic stimulus on transmission is examined in the presence of intact $GABA_A$ receptor-mediated inhibition. In wt animals, a tetanus produces a moderate amount of potentiation. (Fig. 2a) In mutant animals, the potentiation following tetanus is greater than in wt animals. (Fig.. 2a) Differences in potentiation between wt and mutant animals is statistically tested at various time points during a course of under 60 min post-tetanus and is found to be greater in mutants. LTP assessed in the presence of an NMDA receptor antagonist results in

blocking potentiation in both the wt and mutant cells, showing that the mutant cells have enhanced potentiation and potentiation requires NMDA-receptor activation in both types of cells.

Blockade of the $GABA_A$ receptor also differentiates the response between mutant and wt cells. While $GABA_A$ receptor blockade increases LTP in wt animals, there is no significant increase in the mutant cells. It is concluded that the effect of the mutation on potentiation is occluded by blockade of inhibition, indicating that the two factors act on a common pathway.

In another test the effect of an agent on LTP with a moderately strong tetanus, e.g. 3 1sec 100 Hz tetani), the potentiation is larger in mutant as compared to wt animals. However, with agents that increase $GABA_A$ receptor transmission, suppression of the enhanced potentiation should be observed. This can be demonstrated with flunitrazepam as a control or standard with which the effect of the candidate agent may be compared.

Finally, the ratio of peak inhibitory to excitatory responses is significantly greater in mutants as compared to wt. It appears that the observed result is relatively independent of the site of stimulation of the tissue and variations in stimulus intensity. Because of the greater ratio for mutant cells as compared to wt, depending upon the pathway and component of the pathway upon which the agents acts mutants may have a greater response to agents in the reduction of the difference in ratio between mutant and wt cells. This abnormality in mutants (increased inhibitory transmission) is indicated to be a homeostatic (feedback) system that has been turned on in these animals to suppress the underlying aberrant signaling (increased calcium rise). Candidate drugs may not directly affect the inhibitory transmission and still be efficacious, for example, if they act to suppress calcium rise through some other mechanism under the conditions of the assay.

Based on the tests described above, it appears that the presenilin mutation and $GABA_A$ receptor transmission act on the same pathway that regulates potentiation of synaptic transmission. This can be explained by the mutation decreasing $GABA_A$ receptor transmission or the mutation acts downstream of $GABA_A$ receptor transmission, along the signal transduction pathway that generates potentiation. By measuring the effect of an agent on plasticity of mutant cells as compared to wt cells, one may influence the pathway associated with the $GABA_A$ receptor and restore the response toward the wt response.

It is evident from the above results that methods are provided employing mutated mammalian hippocampal cells, conveniently as tissue, which differ from wild-type cells in their increased potentiation as evidenced in their response to tetani. Furthermore, drugs can be screened to determine their effect on returning the response of the mutated cells to a wild-type response. Particularly, a mutation in presenilin protein, which enhances excitability of the cells upon stimuli, allows for screening of drugs which restore wild-type behavior, as demonstrated with a benzodiazepine. By employing tetani under conditions where plasticity of the cells can be determined, an efficient screening tool is provided for determining effectiveness of drugs for the treatment of Alzheimer's disease.

The references described throughout this specification are fully incorporated by reference. Having now fully described the invention, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the invention as set forth herein.

WHAT IS CLAIMED IS:

1. A method for screening for drugs for the treatment of Alzheimer's disease, said method comprising:

contacting mutant hippocampal cells having enhanced synaptic potentiation upon stimulation as compared to wild-type hippocampal cells with a candidate drug;

subjecting said mutant cells to tetanic stimulation; and

determining the effect of said agent on the synaptic potentiation of said mutant hippocampal cells;

wherein a reduction in the enhanced synaptic potentiation of the mutant hippocampal cells is indicative of activity of a candidate drug for the treatment of Alzheimer's disease.

2. A method according to Claim 1, wherein said mutant cells are mutated in a presenilin gene.

3. A method according to Claim 2, wherein said mutant cells are mouse hippocampal tissue slices.

4. A method according to Claim 1, wherein said enhanced synaptic potentiation is as a result of a change in the $GABA_A$ receptor pathway.

5. A method for screening for drugs for the treatment of Alzheimer's disease, said method comprising:

contacting mutant hippocampal cells, having enhanced synaptic potentiation upon stimulation as compared to wild-type hippocampal cells, with a candidate drug;

subjecting said mutant and wild-type hippocampal cells to a tetanic stimulus;

measuring changes in potentiation with time of the mutant and wild-type hippocampal cells and comparing the effect of said agent on the synaptic potentiation of said mutant as compared to the observed synaptic potentiation of said wild-type hippocampal cells;

wherein a reduction in the enhanced synaptic potentiation of the mutant hippocampal cells as compared to the synaptic potentiation of the wild-type cells is indicative of activity of a candidate drug for the treatment of Alzheimer's disease.

6. A method according to Claim 5, including the additional steps of:

contacting mutant hippocampal cells having enhanced synaptic potentiation upon tetanic stimulation as compared to wild-type hippocampal cells with a $GABA_A$ receptor antagonist;

subjecting said mutant and wild-type hippocampal cells to tetanic stimulation; and

measuring changes in synaptic potentiation with time of the mutant and wild-type hippocampal cells and comparing the effect of said $GABA_A$ receptor antagonist on said mutant and said wild-type hippocampal cells;

wherein a reduction in the enhanced synaptic potentiation of the mutant hippocampal cells without a significant change in the synaptic potentiation of the wild-type cells is indicative of the mutation acting on a common pathway with said $GABA_A$ receptor antagonist.

7. A method according to Claim 5, wherein said agent is present with said wild-type hippocampal cells.

8. A method for screening for drugs for the treatment of Alzheimer's disease, said method comprising:

contacting mutant hippocampal cells having enhanced synaptic potentiation upon stimulation as compared to wild-type hippocampal cells with a candidate drug;

subjecting said mutant and wild-type hippocampal cells to a tetanic stimulus at a first potential of glutamate currents and a second potential of $GABA_A$ currents;

measuring the synaptic response at each of the first and second potentials for the mutant and wild-type hippocampal cells and comparing the effect of said agent on said mutant and said wild-type hippocampal cells;

wherein a reduction in the enhanced synaptic response of the mutant hippocampal cells without a significant change in the synaptic response of the wild-type cells is indicative of activity of a candidate drug for the treatment of Alzheimer's disease.

9. A method for screening for drugs for the treatment of Alzheimer's disease, said method comprising:

contacting mutant mouse hippocampal cells mutated in the presenilin-1 gene and having enhanced synaptic potentiation upon tetanic stimulation as compared to wild-type hippocampal cells with a candidate drug;

subjecting said mutant and wild-type hippocampal cells to tetanic stimulation; and

comparing the effect of said agent on said mutant and said wild-type hippocampal cells upon tetanic stimulation;

wherein a reduction in the enhanced synaptic potentiation of the mutant hippocampal cells without a significant change in the synaptic potentiation of the wild-type cells is indicative of activity of a candidate drug for the treatment of Alzheimer's disease.

10. Slices of mouse hippocampal cells having a mutation in a presenilin gene combined with a candidate drug.

11. Slices of mouse hippocampal cells according to Claim 10, after tetanic stimulation.

12. Slices of mouse hippocampal cells according to Claim 10, wherein said mutation is the PS-1 Δ9 mutation.

ABSTRACT OF THE DISCLOSURE

Methods of screening candidate drugs for the treatment of Alzheimer's disease are provided. Employing tissue slices of mutant mouse hippocampal cells having an extra copy of a mutant form of the presenilin-1 gene, it is found that these cells have enhanced potentiation to tetanic stimuli. By subjecting both wild-type and mutant hippocampal cells to tetanic stimuli in the presence of a candidate drug, reduction of the enhanced potentiation of the mutant cells without significant change in the potentiation of the wild-type cells indicates a successful candidate.

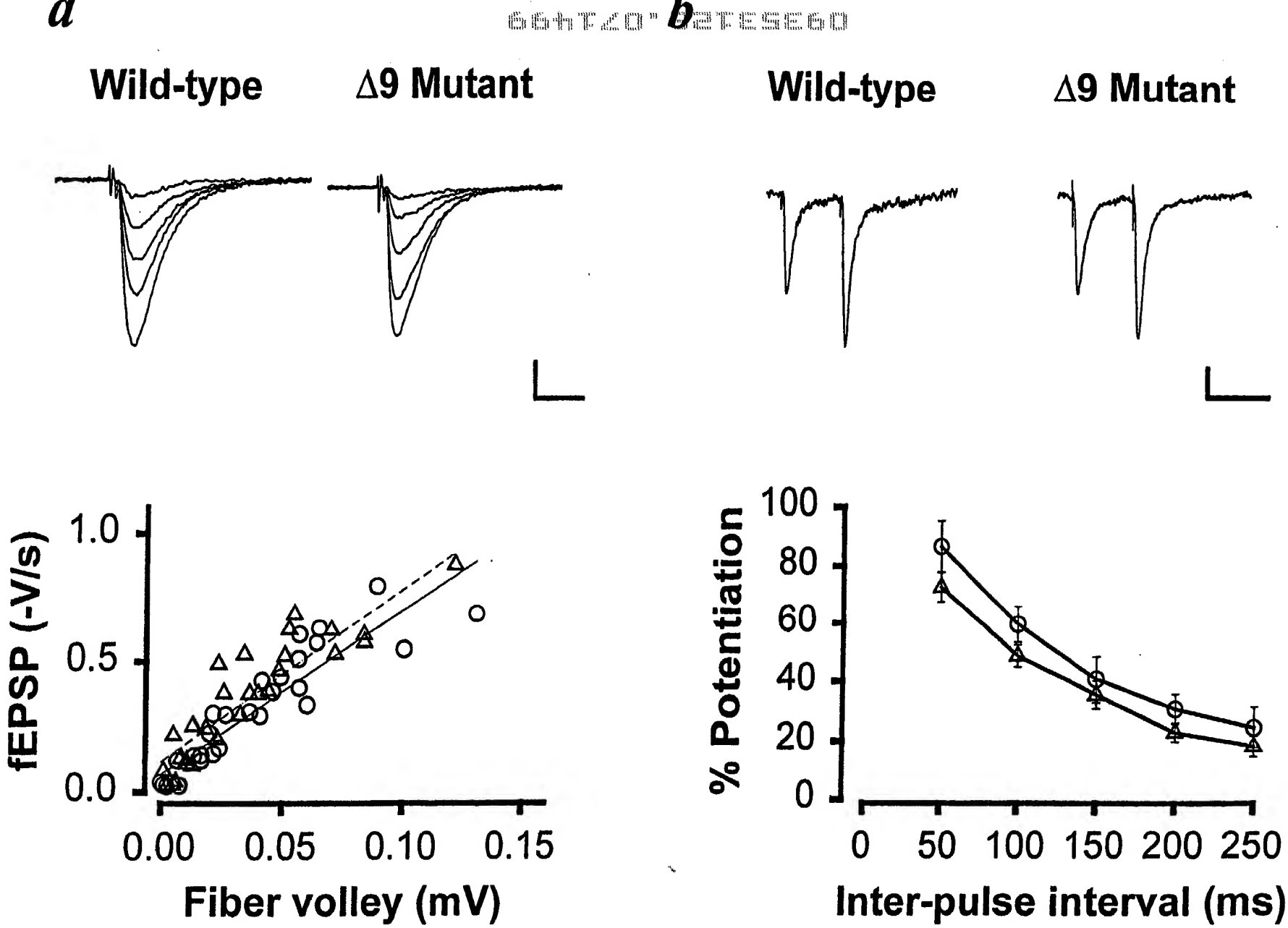

a
b
Wild-type
Δ9 Mutant
Wild-type
Δ9 Mutant
fEPSP (-V/s)
1.0
0.5
0.0
0.00
0.05
0.10
0.15
Fiber volley (mV)
% Potentiation
100
80
60
40
20
0
0
50
100
150
200
250
Inter-pulse interval (ms)

FIGURE 1

FIGURE 2

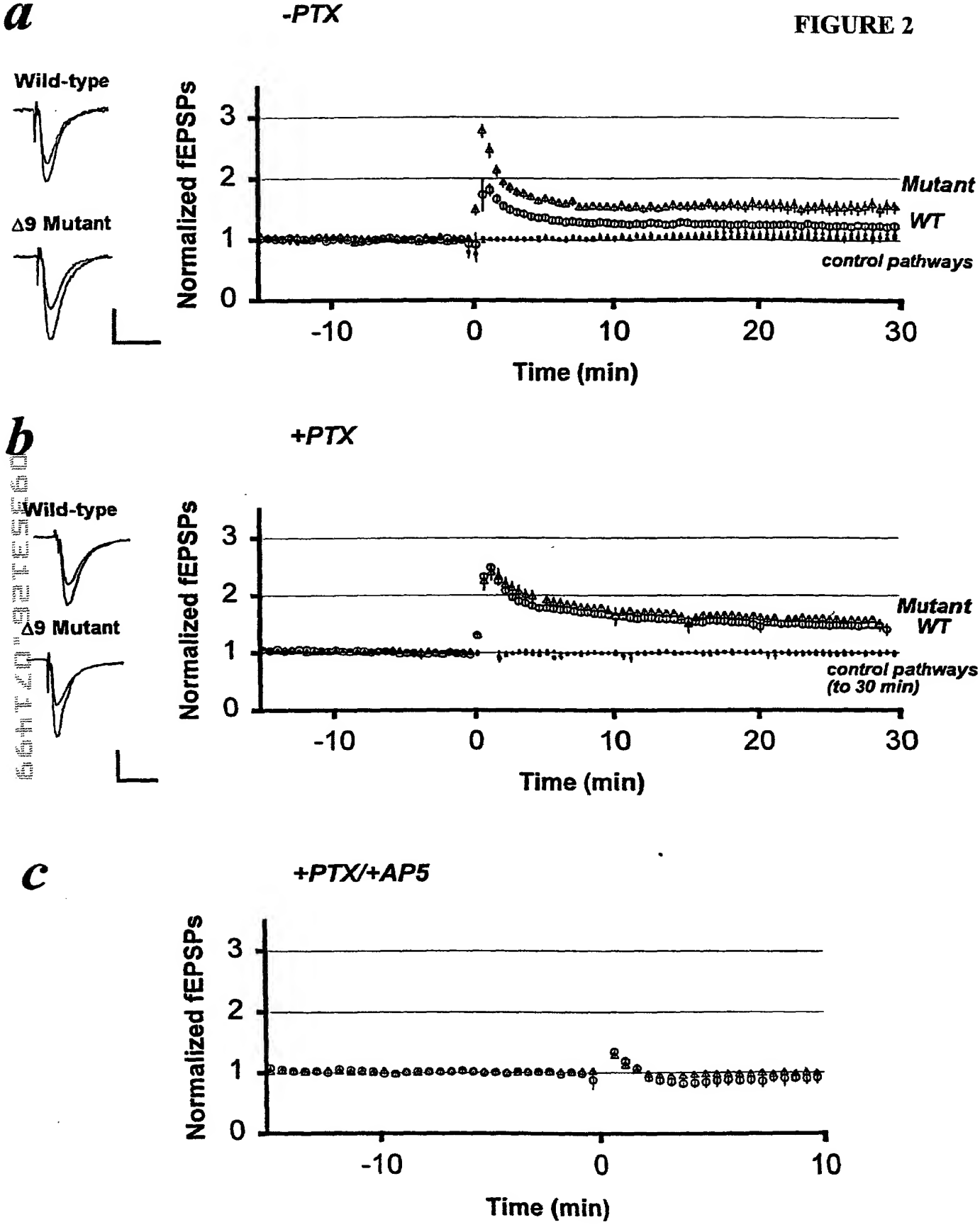
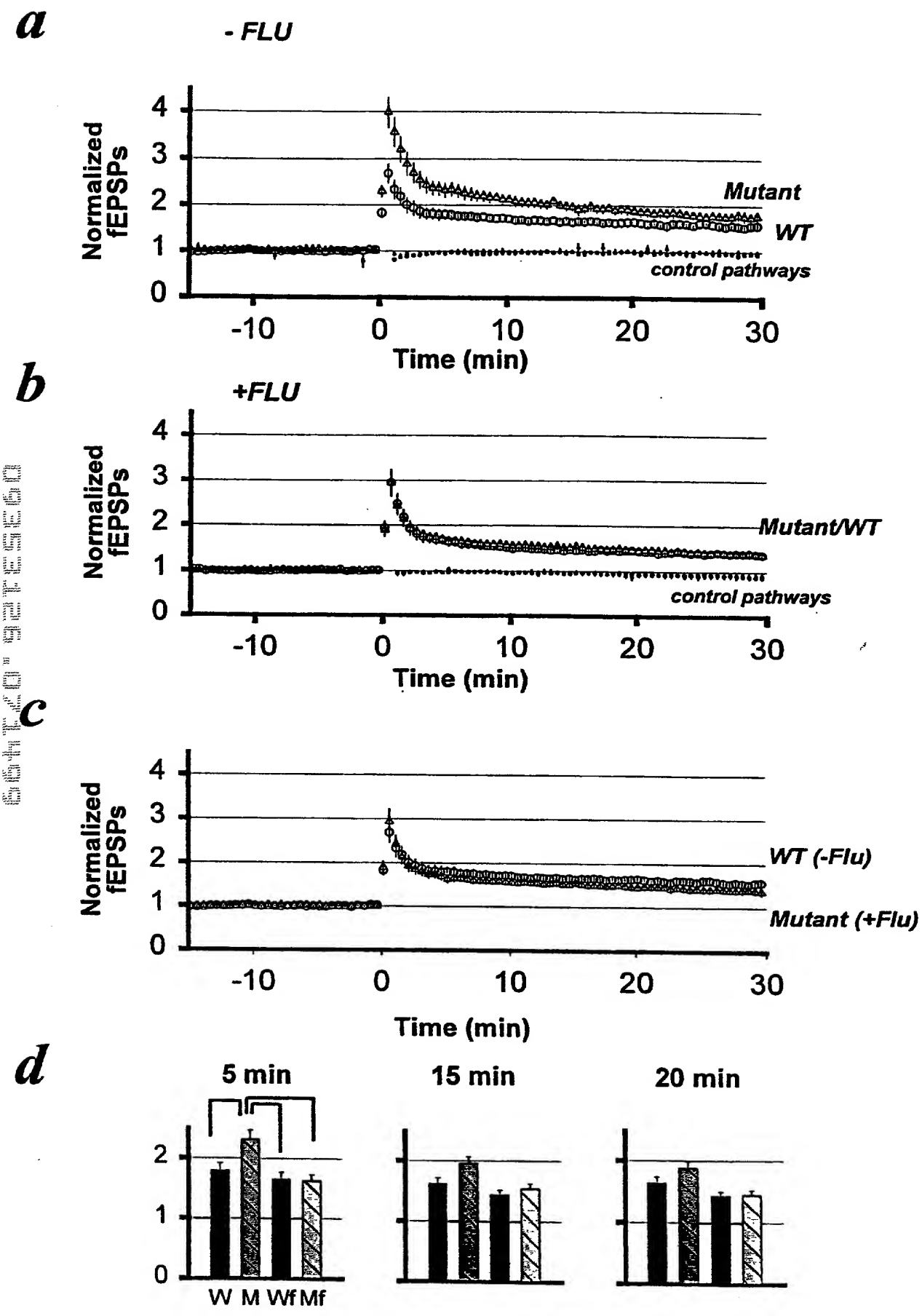

a
-PTX
Wild-type
Δ9 Mutant
Normalized fEPSPs
3
2
1
0
-10
0
10
20
30
Time (min)
Mutant
WT
control pathways
b
+PTX
Wild-type
Δ9 Mutant
Normalized fEPSPs
3
2
1
0
-10
0
10
20
30
Time (min)
Mutant
WT
control pathways
(to 30 min)
c
+PTX/+AP5
Normalized fEPSPs
3
2
1
0
-10
0
10
Time (min)

FIGURE 3


a
- FLU
Normalized fEPSPs
Mutant
WT
control pathways
Time (min)
b
+FLU
Normalized fEPSPs
Mutant/WT
control pathways
Time (min)
c
Normalized fEPSPs
WT (-Flu)
Mutant (+Flu)
Time (min)
d
5 min
15 min
20 min
W M Wf Mf

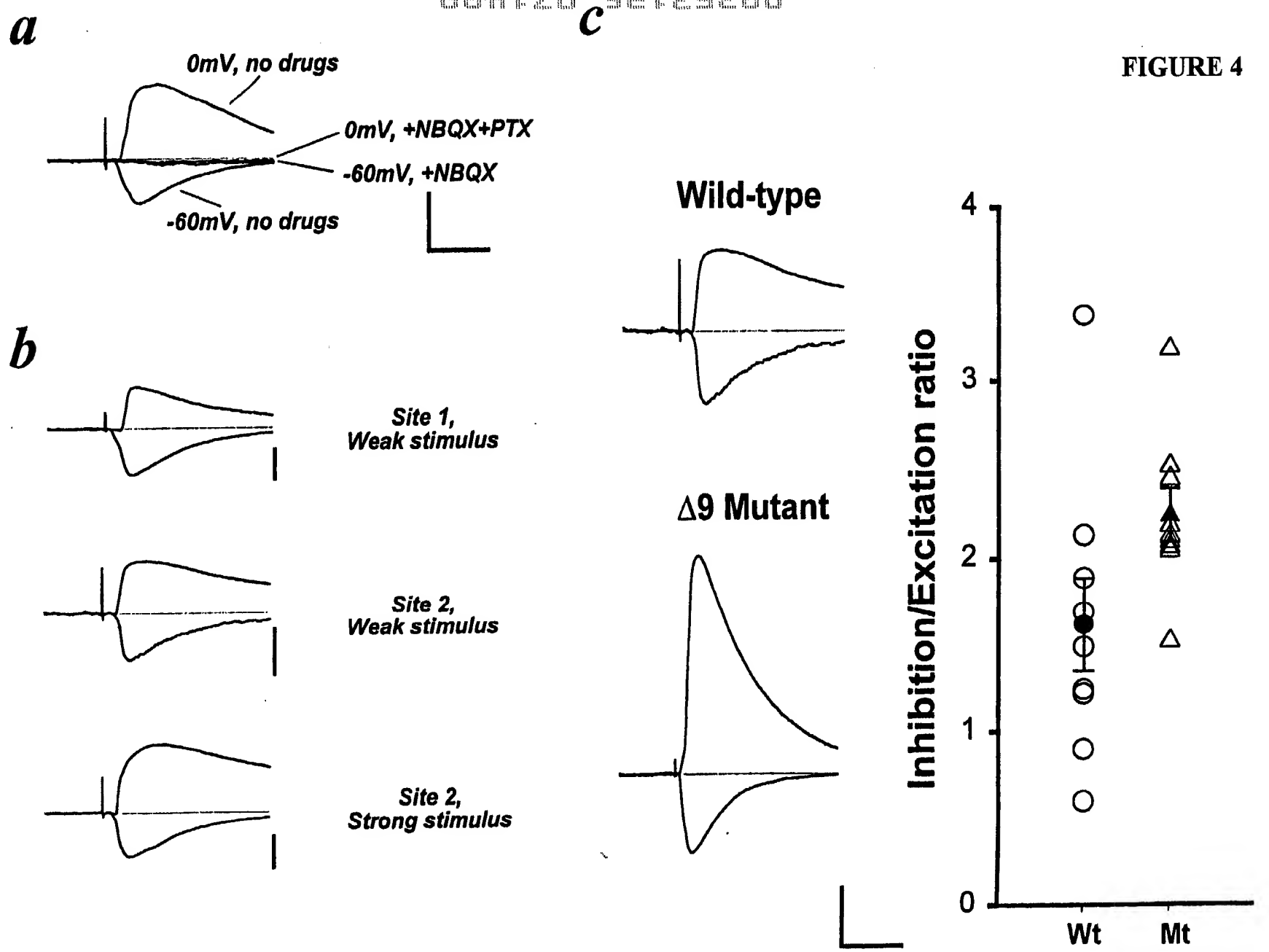


FIGURE 4

PATENT **ATTORNEY DOCKET NO. CSHL.005.00US**

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: **Roberto Malinow** *et al.*	)	
	)	Examiner: Not yet assigned
Serial No.: **09/193,221**	)	
	)	Art Unit: 1614
Filed: **November 16, 1998**	)	
	)	
For: **Diagnostic methods for drug screeing for alzheimer's disease**	)	**REVOCATION AND**
	)	**APPOINTMENT OF NEW POWER**
	)	**OF ATTORNEY**

Commissioner of Patents and Trademarks
Washington, D.C. 20231

Sir:

The undersigned, an authorized representative of Cold Spring Harbor Laboratory, having its principal place of business at One Bungtown Road, Cold Spring Harbor, NY 11724, being assignee of 100% interest of this application, hereby revokes all powers of attorney previously granted in this application and hereby appoints

Barbara Rae-Venter	Reg. No. 32,750
Bertram I. Rowland	Reg. No. 20,015
Ofer Matalon	Reg. No. 39,439

as attorneys with full power of substitution and revocation to prosecute this application and to transact all business in the United States Patent and Trademark Office connected therewith, and hereby request that all correspondence regarding this application be sent to the firm of:

CERTIFICATE OF FIRST-CLASS MAILING

I hereby certify that this paper or fee is being deposited with the United States Postal Service as first-class mail in an envelope addressed to the Commissioner of Patents and Trademarks, Washington, D.C. 20231 on 3-3-99.
(Date)

(Signature)

Janine Fernandez
(Printed Name)

Rae-Venter Law Group, P.C.
P. O. Box 60039
Palo Alto, CA 94306

Telephone No.: 650-328-4400
Facsimile No.: 650-328-4477

The undersigned has reviewed the chain of title and to the best of the undersigned's knowledge, title is in the assignee identified above.

Date: 2/24/99 By: [signature]

Title: John Maroney
Assistant Administrative Director

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Palo Alto, CA 94306
Telephone No. 650-328-4400
Facsimile No. 650-328-4477

BRV/mnb

POWER OF ATTORNEY BY ASSIGNEE
(Not Accompanying Application)

The undersigned assignee of the entire interest in application for letters patent entitled: **DIAGNOSTIC METHODS FOR DRUG SCREENING FOR ALZHEIMER'S DISEASE** and having the named inventors: Roberto Malinow, Shahin Zaman, Sangram S. Sisodia, David R. Borchelt, Michael K. Lee, bearing Serial No. 09/193,221 filed November 16, 1998 hereby appoints the following attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith; said appointment to be to the exclusion of the inventor(s) and his (their) attorney(s) in accordance with the provisions of 37 C.F.R. 1.32: Harold C. Hohbach, Reg. No. 17,757; Aldo J. Test, Reg. No. 18,048; Thomas O. Herbert, Reg. No. 18,612; Bertram I. Rowland, Reg. No. 20,015; Donald N. MacIntosh, Reg. No. 20,316; Jerry G. Wright, Reg. No. 20,165; Edward S. Wright, Reg. No. 24,903; David J. Brezner, Reg. No. 24,774; Richard E. Backus, Reg. No. 22,701; James A. Sheridan, Reg. No. 25,435; Robert B. Chickering, Reg. No. 24,286; Gary S. Williams, Reg. No. 31,066; Richard F. Trecartin, Reg. No. 31,801; Walter H. Dreger, Reg. No. 24,190; Steven F. Caserza, Reg. No. 29,780; William S. Galliani, Reg. No. 33,885; Laura L. Kulhanjian, Reg. No. 33,257; Julian Caplan, Reg. No. 14,785; R. Michael Ananian, Reg. No.35, 050; provided that if any one of said attorneys ceases being affiliated with the law firm of Flehr Hohbach Test Albritton & Herbert LLP as partner, employee or of counsel, such attorney's appointment as attorney and all powers derived therefrom shall terminate on the date such attorney ceases being so affiliated.

In accordance with 37 CFR 3.73 the assignee hereby certifies that the evidentiary documents with respect to its ownership have been reviewed and that, to the best of assignee's knowledge and belief, title is in the assignee seeking to take this action.

Direct all telephone calls to BERTRAM I. ROWLAND at (650) 781-1989.

Address all correspondence to:

FLEHR HOHBACH TEST
ALBRITTON & HERBERT LLP
Suite 3400, Four Embarcadero Center
San Francisco, California 94111

DATED: December 3, 1998

COLD SPRING HARBOR LABORATORY

By: John Maroney
(typed name)

Signature: [signature]

Title: John Maroney
~~Assistant Administrative Director~~

Address: 1 BUNGTOWN ROAD

COLD SPRING HARBOR, NEW YORK 11724

File No. A-67162/BIR

Form #1.09 (8058)
04/95

DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

the specification of which

(check one) ☐ is attached hereto.

☒ was filed on November 16, 1998 as Application Serial No. 09/193,221 and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the Patent Office all information known to me to be material to patentability as defined in 37 C.F.R. 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed	
			☐	☐
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No
			☐	☐
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No
			☐	☐
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose to the Patent Office all information known to me to be material to patentability as defined in 37 C.F.R. 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)

I hereby appoint the following attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: Harold C. Hohbach, Reg. No. 17,757; Aldo J. Test, Reg. No. 18,048; Thomas O. Herbert, Reg. No. 18,612; Bertram I. Rowland, Reg. No. 20,015; Donald N. MacIntosh, Reg. No. 20,316; Jerry G. Wright, Reg. No. 20,165; Edward S. Wright, Reg. No. 24,903; David J. Brezner, Reg. No. 24,774; Richard E. Backus, Reg. No. 22,701; James A. Sheridan, Reg. No. 25,435; Robert B. Chickering, Reg. No. 24,286; Gary S. Williams, Reg. No. 31,066; Richard F. Trecartin, Reg. No. 31,801; Walter H. Dreger, Reg. No. 24,190; Steven F. Caserza, Reg. No. 29,780; William S. Galliani, Reg. No. 33,885; Laura L. Kulhanjian, Reg. No. 33,257; Julian Caplan, 14,785; R. Michael Ananian, Reg. No. 35,050; provided that if any one of said attorneys ceases being affiliated with the law firm of Fleh, Hohbach Test Albritton & Herbert LLP as partner, employee or of counsel, such attorney's appointment as attorney and all powers derived therefrom shall terminate on the date such attorney ceases being so affiliated.

Direct all telephone calls to BERTRAM I. ROWLAND at (415) 781-1989.

Address all correspondence to:

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ALBRITTON & HERBERT LLP
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San Francisco, California 94111

File No. A-67162/BIR

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Title 18, United States Code, §1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor: Roberto Malinow

Inventor's signature: [signature]

Date: 12.4.98

Residence Address: 1 Bennington Rd
Cold Spring Harb NY 11724

Citizenship: UNITED STATES

Full name of second joint inventor: Shahid Zaman

Inventor's signature: _____

Date: _____

Residence Address: _____

_____

Citizenship: UNITED KINGDOM

I hereby appoint the following attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: Harold C. Hohbach, Reg. No. 17,757; Aldo J. Test, Reg. No. 18,048; Thomas O. Herbert, Reg. No. 18,612; Bertram I. Rowland, Reg. No. 20,015; Donald N. MacIntosh, Reg. No. 20,316; Jerry G. Wright, Reg. No. 20,165; Edward S. Wright, Reg. No. 24,903; David J. Brezner, Reg. No. 24,774; Richard E. Backus, Reg. No. 22,701; James A. Sheridan, Reg. No. 25,435; Robert B. Chickering, Reg. No. 24,286; Gary S. Williams, Reg. No. 31,066; Richard F. Trecartin, Reg. No. 31,801; Walter H. Dreger, Reg. No. 24,190; Steven F. Caserza, Reg. No. 29,780; William S. Galliani, Reg. No. 33,885; Laura L. Kulhanjian, Reg. No. 33,257; Julian Caplan, 14,785; R. Michael Ananian, Reg. No. 35,050; provided that if any one of said attorneys ceases being affiliated with the law firm of Fleh, Hohbach Test Albritton & Herbert LLP as partner, employee or of counsel, such attorney's appointment as attorney and all powers derived therefrom shall terminate on the date such attorney ceases being so affiliated.

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San Francisco, California 94111

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Full name of sole or first inventor: Roberto Malinow

Inventor's signature: _____

Date: _____

Residence Address _____

_____

Citizenship: UNITED STATES

Full name of second joint inventor: Shahid Zaman

Inventor's signature: [signature]

Date: 4th Jan. 1999

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Citizenship: UNITED KINGDOM

Full name of third joint inventor: Sangram S. Sisodia

Inventor's signature: [signature]

Date: December 3, 1998

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Citizenship: UNITED STATES

Full name of fourth joint inventor: David R. Borchelt

Inventor's signature:

Date:

Residence Address:

Citizenship: UNITED STATES

Full name of fifth joint inventor: Michael K. Lee

Inventor's signature:

Date:

Residence Address:

Citizenship: UNITED STATES

Full name of third joint inventor: Sangram S. Sisodia

Inventor's signature:

Date:

Residence Address

Citizenship: UNITED STATES

Full name of fourth joint inventor: David R. Borchelt

Inventor's signature: David R. Borchelt

Date: 12-14-98

Residence Address: 723 Stevenson Ln
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Citizenship: UNITED STATES

Full name of fifth joint inventor: Michael K. Lee

Inventor's signature:

Date:

Residence Address

Citizenship: UNITED STATES

Full name of third joint inventor: Sangram S. Sisodia

Inventor's signature:

Date:

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Citizenship: UNITED STATES

Full name of fourth joint inventor: David R. Borchelt

Inventor's signature:

Date:

Residence Address

Citizenship: UNITED STATES

Full name of fifth joint inventor: Michael K. Lee

Inventor's signature: [signature]

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